Rhein: an anthraquinone that modulates superoxide anion production from human neutrophils

M. MIAN, S. BRUNELLESCHI^{*}, S. TARLI, A. RUBINO, D. BENETTI[†], R. FANTOZZI[‡], L. ZILLETTI[†], Department of Preclinical and Clinical Pharmacology, University of Florence, V. le Morgagni, 65-50134 Firenze, [†]Istituto Gentili S.p.A., Pisa, [‡]Institute of Pharmacology, University of Ferrara, Ferrara, Italy

Rhein (4,5-dihydroxyanthraquinone-2-carboxylic acid), the active metabolite of diacetylrhein, which has been reported as an effective antirheumatic drug in man, inhibited superoxide anion production from human neutrophils challenged with *N*-formylmethionyl-leucyl-phenylalanine (FMLP: IC50, 2×10^{-5} M) and A23186 (IC50, 10^{-5} M), but not with phorbol myristate acetate. In the same concentration range $(10^{-6}-10^{-3}$ M), the drug did not affect oxy-radical production by a cell-free hypoxanthine-xanthine oxidase system and exerted weak inhibitory effects on FMLP-evoked lysosomal enzyme release. Rhein inhibitory effects on neutrophil functioning may contribute to the overall therapeutic activity of the parent drug, diacetylrhein.

Rhein (4,5-dihydroxyanthraquinone-2-carboxylic acid), an anthraquinone from plants of the genus Cassia, has been described as having weak antibacterial effects and cathartic properties (Kean 1970). It is a derivative of sennosides and these anthraquinones have been demonstrated to stimulate colonic fluid and electrolyte secretion and PGE₂ formation in the rat colon, in-vivo (Beubler & Kollar 1985).

The drug was reported to interfere with mitochondrial electron transport and to inhibit flavoproteins concerned with redox reactions of pyridine nucleotides (Kean 1968, 1970; Kean et al 1971). Furthermore, Raimondi et al (1982) showed that rhein, in the concentration range 10^{-5} - 10^{-4} M, inhibited some proteases in-vitro, with bovine pancreatic carboxypeptidase A and porcine pancreatic elastase being the most sensitive enzymes.

Rhein is the active metabolite of diacetylrhein (DAR: 1,8-diacetoxy-9, 10-dioxo-dihydroanthracene-3-carboxylic acid), which some authors have reported to exert anti-inflammatory effects in in-vivo animal tests and to display antirheumatic activity in man (Kay et al 1980; Neuman 1980; Pomarelli et al 1980). Diacetylrhein, in in-vivo experiments, and rhein, in in-vitro experiments, did not inhibit prostaglandin synthesis in inflammatory exudates and in guinea-pig lung homogenates that had been incubated with arachidonic acid and challenged with rhein in the concentration range $3\cdot 2 \times 10^{-6} - 3\cdot 2 \times 10^{-4}$ M (Pomarelli et al 1980; Franchi-Micheli et al 1983).

Human neutrophils, a cell type that plays a key role in inflammatory processes, are known to degranulate.

aggregate and experience a 'respiratory burst' (which is characterized by the generation of reactive oxygenderived free radicals), in response to a variety of soluble and insoluble agents. An NAD(P)H-dependent flavoprotein has been implicated in superoxide anion (O_2^{-}) production from human neutrophils (Babior 1984), whereas tissue destruction in chronic disorders (such as emphysema and arthritis) has been related to the release of lysosomal proteases, and particularly to the release of the azurophil granule enzyme, elastase (Saklatvala 1977; Werb et al 1982). Because of the efficacy of DAR in-vivo and rhein's ability to inhibit protease enzymes and flavoproteins in-vitro, we decided to verify whether or not it could interfere with O2- production and lysosomal enzyme release from human neutrophils.

The effects of rhein were evaluated in cells which had been activated in-vitro by different stimuli: a chemotactic peptide (*N*-formylmethionyl-leucyl-phenylalanine: FMLP), a calcium ionophore (A23187) and an activator of protein-kinase C (phorbol myristate acetate, PMA: Nishizuka 1984).

Materials and methods

Neutrophils were isolated from healthy adult volunteers by standard techniques of dextran sedimentation (dextran T500, Pharmacia), Ficoll-Paque (Pharmacia) gradient centrifugation and hypotonic lysis of erythrocytes, as previously described (Fantozzi et al 1986).

Neutrophils were suspended in a buffered salt solution (mM: NaCl 138, KCl 2·7, Na₂HPO₄ 8·1, KH₂PO₄ 1·5, MgCl₂ 1, CaCl₂ 2, pH 7·4) supplemented with glucose (1 mg mL⁻¹) and human serum albumin (1 mg mL⁻¹: Behringwerke). When indicated, experiments were carried out by using a calcium-free medium containing EGTA [1 mM; ethylene glycol-bis (β -amino-ethyl ether) N, N, N', N'-tetraacetic acid: Sigma].

Cells were treated with cytochalasin B ($5 \mu g m L^{-1}$; Aldrich) for 5 min, before exposure to rhein (kindly supplied by Istituto Gentili S.p.A., Pisa, Italy, as sodium salt). After incubation with rhein, neutrophils were challenged with different stimuli (FMLP: Serva; A23187: Calbiochem; PMA: Sigma) at 37 °C.

 $O_2^{-\tau}$ production was continuously monitored spectrophotometrically at 37 °C, by determining superoxide dismutase (SOD: Boehringer-Mannheim)-inhibitable

^{*} Correspondence.

cytochrome C (Boehringer-Mannheim) reduction as previously described (Fantozzi et al 1986) and expressed as nmoles cytochrome C reduced $\min^{-1}/10^6$ cells (Smolen et al 1981).

β-Glucuronidase (substrate: phenolphthalein glucuronic acid, Sigma) and myeloperoxidase (substrate: *o*-dianisidine; Sigma) were determined as reported by Fantozzi et al (1986). Elastase (substrate: methoxysuccinyl-L-alanyl-L-prolyl-L-valine *p*nitroanilide; Sigma) was measured according to Hojima et al (1983).

Xanthine oxidase (XOD) and hypoxanthine (HPX) were from Boehringer-Mannheim and Sigma, respectively.

Rhein inhibitory effects were expressed as percentage inhibition of the values obtained with the stimulus alone.

Results

Rhein, in the concentration range 10^{-6} – 10^{-3} M, inhibited O_2^{-7} production from FMLP-activated human neutrophils: the effect was dose-dependent (IC50 = 2 × 10^{-5} M) and maximal inhibition was achieved at 1.7×10^{-4} M rhein (Fig. 1A).

In the experiments depicted in Fig. 1A, neutrophils were preincubated with rhein for 10 min; however, O_2 production was inhibited to the same extent even when cells had been treated with the drug for different times (up to 30 min). Similar inhibitory effects were recorded by adding rhein immediately before the stimulus.

Rhein, even at the highest concentration $(3.5 \times 10^{-4} \text{ M})$, did not affect oxy-radical production by a cell-free XOD $(0.2 \text{ U mL}^{-1})/\text{HPX} (10^{-4} \text{ M})$ system.

When the experiments were performed by incubating human neutrophils in a calcium-free medium containing EGTA 1 mM, rhein still affected O_2^{-r} production from FMLP-activated neutrophils. The effect was dosedependent and inhibition was similar to that recorded in the presence of calcium in the medium (Fig. 1B).

Rhein inhibited O₂- production from A23187-stimulated neutrophils in a dose-dependent manner (IC50 = 10^{-5} M). Maximal inhibitory effects were similar to those obtained in FMLP-activated cells (Fig. 2). When neutrophils were challenged with PMA, rhein was devoid of any inhibitory action, even at the concentration of 3.5×10^{-4} M, which exerted maximal effects on O₂- production from FMLP- and A23187-activated human neutrophils (Fig. 2). In the same concentration range, rhein not only inhibited O2- production, but also affected lysosomal enzyme (e.g., β-glucuronidase, elastase, myeloperoxidase) release from FMLP-stimulated cells. The extent of inhibition was significantly less than that recorded by evaluating O_2^{-} production; a 30% inhibition of β -glucuronidase, myeloperoxidase and elastase release was obtained at the rhein concentration of 7×10^{-5} M.

Higher concentrations of rhein directly inhibited the enzyme activities that were evaluated in the super-



FIG. 1. Effect of rhein on FMLP-evoked superoxide anion production from human neutrophils. (A) Cells stimulated by FMLP 10^{-7} m produced 4.28 ± 0.5 nmol cytochrome C reduced min⁻¹/10⁶ cells (n =7). Values obtained in the absence of rhein were taken as 100%. Results are the means \pm s.e.m. of 7 experiments. (B) \Box : cells suspended in a medium containing 1 mm calcium produced $5.37 \pm$ 0·1 nmol cytochrome C reduced min⁻¹/10⁶ cells. \blacksquare : cells incubated in a calcium-free medium plus 1 mm EGTA produced 2·26 \pm 0·1 nmol cytochrome C reduced min⁻¹/ 10⁶ cells. Values are the means \pm s.e.m. of 3 experiments.

natants of FMLP-challenged human neutrophil suspensions: a 50% inhibition of elastase activity was measured at the rhein concentration of 3.5×10^{-4} M.

Discussion

The results presented here demonstrate that rhein inhibited O_2^{-} production from human neutrophils, without exerting scavenging effects in a cell-free XOD/ HPX system.

Rhein displayed a stimulus-selectivity, since it inhibited with similar potencies O_2^- production from human neutrophils challenged with FMLP or A23187, but not with PMA. The reasons for this stimulus-selectivity are not clear at present, but might be related to the different abilities of these agents to affect calcium-dependent steps that underlie neutrophil activation, since calcium is widely believed to represent a major mediator of neutrophil activation (Naccache et al 1985; Westwick & Poll 1986).



Fig. 2. Effect of rhein on superoxide anion production from human neutrophils challenged with A23187 or PMA. Cells stimulated by A23187 10^{-5} M (\Box) produced 1.63 ± 0.1 nmol cytochrome C reduced min⁻¹/10⁶ cells (n = 3). Cells challenged with PMA 50 µg mL⁻¹ (\blacksquare) produced 9.43 ± 0.2 nmol cytochrome C reduced min⁻¹/10⁶ cells (n = 3). Values obtained in the absence of rhein were taken as 100%. Results are the means ± s.e.m. of 3 experiments.

Actually, FMLP was capable of evoking O_2^{-} production even in the presence of EGTA, while A23187 greatly depended on extracellular calcium to induce O_2^{-} production (Pozzan et al 1983; Smolen et al 1981; Sullivan et al 1984; Brunelleschi et al, unpublished observations); PMA, which was suggested to mimic the effects of diacylglycerol, stimulated O_2^{-} production without inducing a rise in cytosolic calcium (Nishizuka 1984; Westwick & Poll 1986). Rhein was suggested to possess potential chelating properties for calcium (Kean 1968). In our experiments, the drug showed inhibitory effects on FMLP-stimulated neutrophils, even when the cells were exposed to EGTA in a calcium-free medium.

The ability of rhein to interfere with O_2^{-} production from human neutrophils could depend, at least partially, on its inhibitory effects on oxidase systems, since the drug was reported to inhibit competitively NADHdehydrogenase, in-vitro (Kean et al 1971). Neutrophils require a NAD(P)H-dependent membrane oxidase for generating O_2^{-} (Babior 1984).

Rhein slightly inhibited lysosomal enzyme release from FMLP-activated neutrophils and, at the highest concentrations, directly affected the activities of the enzymes that were released. These data have to be evaluated along with those obtained by Raimondi et al (1982). The capacity of some protease inhibitors (e.g., L-1-tosylamide-2-phenylethylchloromethylketone : TPCK; aprotinin) to inhibit oxy-radical production from human neutrophils was reported (Kitagawa et al 1980; Curnutte et al 1984; Hallett et al 1985).

In conclusion, rhein appeared to affect respiratory burst from human neutrophils and, to a lesser extent, lysosomal enzyme release. These inhibitory effects might account for some of the in-vivo pharmacological activities of its precursor, diacetylrhein.

REFERENCES

- Babior, B. M. (1984) J. Clin. Invest. 73: 599-601
- Beubler, E., Kollar, G. (1985) J. Pharm. Pharmacol. 37: 248-251
- Curnutte, J. T., Badwey, J. A., Robinson, J. M., Karnovsky, M. J., Karnovsky, M. L. (1984) J. Biol. Chem. 259: 11851–11857
- Fantozzi, R., Brunelleschi, S., Rubino, A., Tarli, S., Masini, E., Mannaioni, P. F. (1986) Agents Actions 18: 155–158
- Franchi-Micheli, S., Lavacchi, L., Friedmann, C. A., Zilletti, L. (1983) J. Pharm. Pharmacol. 35: 262–264
- Hallett, M. B., Shandall, A., Young, H. L. (1985) Biochem. Pharmacol. 34: 1757-1761
- Hojima, Y., Pisano, J. J., Cochrane, C. G. (1983) Ibid. 32: 985–990
- Kay, A. G. L., Griffiths, L. G., Volans, G. N., Grahame, R. (1980) Curr. Med. Res. Opin. 6: 548–551
- Kean, E. A. (1968) Arch. Biochem. Biophys. 127: 528-533
- Kean, E. A. (1970) Biochem. Pharmacol. 19: 2201-2210
- Kean, E. A., Gutman, M., Singer, T. P. (1971) J. Biol. Chem. 246: 2346–2353
- Kitagawa, S., Takaku, F., Sakamoto, S. (1980) J. Clin. Invest. 65: 74–81
- Naccache, P. H., Molski, T. F. P., Borgeat, P., White, J. R., Sha'afi, R. I. (1985) J. Biol. Chem. 260: 2125–2131
- Neuman, M. (1980) Drug Exp. Clin. Res. 6: 53-64
- Nishizuka, Y. (1984) Nature 308: 693-698
- Pomarelli, P., Berti, M., Gatti, M. T., Mosconi, P. (1980) Il Farmaco Ed. Sci. 35: 836-842
- Pozzan, T., Lew, D. P., Wollheim, C. B., Tsien, R. Y. (1983) Science 221: 1413–1415
- Raimondi, L., Banchelli Soldaini, G., Buffoni, F., Ignesti, G., Massacesi, L., Amaducci, L., Friedmann, C. A. (1982) Pharmacol. Res. Commun. 14: 103–112
- Saklatvala, J. (1977) J. Clin. Invest. 59: 794-801
- Smolen, J. E., Korchak, H. M., Weissmann, G. (1981) Biochim. Biophys. Acta 677: 512-520
- Sullivan, G. W., Donowitz, G. R., Sullivan, J. A., Mandell, G. L. (1984) Blood 64: 1184–1192
- Werb, Z., Banda, M. J., McKerrow, J. H., Sandhaus, R. A. (1982) J. Invest. Dermatol. 79: 154S–159S
- Westwick, J., Poll, C. (1986) Agents Actions 19: 80-86